**The Claustrum of the Sheep and its Connections to the Visual Cortex.**

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**Conflict of Interest**

The authors declare no conflict of interest.

**Data Availability Statement**

The data that support the findings of this study are available from the corresponding author upon reasonable request.

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**ABSTRACT**

The present study analyses the organization and selected neurochemical features of the claustrum and visual cortex of the sheep, based on the patterns of calcium-binding proteins expression. Connections of the claustrum with the visual cortex have been studied by tractography.

Parvalbumin-immunoreactive (PV-ir) and Calbindin-immunoreactive (CB-ir) cell bodies increased along the rostro-caudal axis of the nucleus. Calretinin (CR)-labeled somata were few and evenly distributed along the rostro-caudal axis. PV and CB distribution in the visual cortex was characterized by larger round and multipolar cells for PV, and more bitufted neurons for CB. The staining pattern for PV was the opposite of that of CR, which showed densely stained but rare cell bodies.

Tractography shows the existence of connections with the caudal visual cortex. However, we detected no contralateral projection in the visuo-claustral interconnections. Since sheep and goats have laterally placed eyes and a limited binocular vision, the absence of contralateral projections could be of prime importance if confirmed by other studies, to rule out the role of the claustrum in stereopsis.

**INTRODUCTION**

The claustrum (Cl) is a subcortical structure, relaying inputs from, to, and among cortical areas (Day-Brown et al. 2016; Reser et al., 2017; Wang et al., 2017; White & Mathur, 2018; Krimmel et al., 2019; Jackson et al., 2020), including a rather large connection to and from visual areas (Norita, 1977; Riche and Lanoir, 1978; Olson and Graybiel, 1980; LeVay and Sherk, 1981; Maçarico da Costa et al., 2010). Other data suggest also inputs from serotonergic raphe nuclei (Baizer, 2001), thalamic nuclei (LeVay and Sherk, 1981; Carey and Neal, 1986; Vertes and Hoover, 2008), endopiriform nucleus (Lipowska et al., 2000) and dopaminergic neurons of the ventral tegmental area or *substantia nigra* (Pirone et al., 2018).

Despite the wealth of data from various species on general anatomy, cytoarchitecture and chemoarchitecture, the structure, function and origin of the Cl are still a matter of debate (Edelstein & Denaro, 2004; Crick & Koch, 2005; Pirone et al. 2012; Hinova-Palova et al., 2014a; Hinova-Palova et al., 2014b; Mathur, 2014; Deutch & Mathur, 2015; Goll et al. 2015; Binks et al., 2019; Hinova-Palova et al., 2019a; Hinova-Palova et al., 2019b; Pirone et al., 2020). To this effect, the ontology of the claustro-insular complex is still subject to debate (for in-depth discussion see Butler et al., 2011; Pirone et al., 2012), but homologies reaching birds and even reptiles have been put forward (Puelles et al., 2016, Watson and Puelles, 2017).

An important step in understanding the Cl function is to clarify the neurochemistry of the nucleus and identify its reciprocal connections with other brain areas. Efforts are now increasingly drawn towards precising these connections. As an example, here we mention that there has been no knowledge of direct connection of the Cl to the non-visual thalamus (Carey & Neal, 1986; Day-Brown et al., 2016), until fairly recently (Atlan et al, 2018; Narikiyo et al., 2020).

The Cl contains several types of interneurons, characterized neurochemically by the presence of calcium binding proteins (CBPs), nNOS, the synthetic enzyme for nitric oxide, and different neuropeptides (Eiden et al., 1990; Hinova-Palova et al., 2008; Kowiański et al., 2008; Pirone et al., 2014; Hinova-Palova et al., 2012; Cozzi et al., 2014; Landzhov et al., 2017). On the contrary, the presence of projecting neurons in the Cl is still open. It is well-known that CBPs are classically expressed by GABA-ergic interneurons, but they have also been found in projecting neurons of different brain structures (Gerfen et al., 1985; Bennett-Clarke et al., 1992; Rausell et al., 1992; Liu et al., 2014; Shang et al., 2019; Baizer et al., 2011; Celio, 1990). This implies that some of the CBP-containing neurons in the Cl may indeed project to other brain structures.

The visual cortex of the sheep has been relatively well studied compared to other ungulates. Rose (1942) found that the striated cortex stretched along most of the lateral and entolateral sulcus. Later electrophysiological studies pointed out to a much smaller dorso-occipital part of the sheep cortex (Clarke and Whitteridge, 1976, Clarke, et al., 1976), whereas thalamo-cortical tracing studies confirmed a wider visual territory (Karamanlidis et al., 1979).

One of the now often used investigation techniques involves the mapping of probabilistic tracts using diffusion tensor imaging (DTI), which has recently led a group to push for the establishment of a claustro-cortical fan (Fernàndez-Miranda et al., 2008). In the present study we focused on the connections of the Cl with the primary visual cortex (V1), using the sheep, a large-brained mammals often used in translational research, as experimental model, and a combination of DTI and neurochemistry as working methodology.

**MATERIAL AND METHODS**

**Animals and tissue sampling**

For the present study we utilized the brains of six adult sheep collected at a local slaughterhouse (Table 1). Animals were treated according to the European Community Council directive (86/609/EEC) concerning animal welfare during the commercial slaughtering process and were constantly monitored under mandatory official veterinary medical care.

The ovine brains were extracted within 15 minutes after death, and cut into transverse blocks (0.5 cm thick) containing the CL and the adjoining structures in their rostro-caudal extent. Blocks from the right hemisphere were fixed by immersion in 4% paraformaldehyde in 0.1 M phosphate-buffered saline at pH 7.4 (PBS) and later processed for paraffin embedding. Simultaneously, blocks of the left primary visual cortex were sampled across the splenial and endolateral sulcus.

**Table 1.** specimen data

|  |  |  |  |
| --- | --- | --- | --- |
| Age | Weight | Sex | Breed |
| Adult (4 years) | 34 | F | Brogna |
| Adult (4 years) | 47 | F | Brogna |
| Adult (4 years) | 35.5 | F | Brogna |
| Adult (4 years) | 43,5 | F | Brogna |
| Adult (4 years) | 38 | F | Brogna |
| Adult (4 years) | 41 | F | Brogna |

**Histology and Immunohistochemistry**

MRI images and coronal blocks of sheep brain, later sectioned and stained with cresyl-violet, were used to determine the shape and extent of the Cl.

Immunoperoxidase reaction was performed on serial paraffin sections (5 μm) from four brain blocks representing the Cl rostro-caudal extent (fig. 1). Immunoreaction was carried out employing the following antibodies against three CBPs: a mouse monoclonal anti-parvalbumin (PV, 1:2000, Sigma), a mouse monoclonal anti-parvalbumin (PV, 1:2000, Swant) a mouse monoclonal anti-calbindin D-28K (CB, 1:2000, Sigma), a rabbit polyclonal anti-calbindin D-28k (CB, 1:1000, Swant), a rabbit polyclonal anti-calretinin (CR, 1:100, Abcam), and a rabbit anti-calretinin (CR, 1:1000, Swant) (details are reported in Table 2). Epitope retrieval was carried out at 120 °C in a pressure cooker for 3 min with a Tris/EDTA buffer, pH 9.0. Sections were pretreated with 1% H2O2 in PBS, for 10 min to quench endogenous peroxidase activity, then rinsed with 0.05% Triton-X (TX)-100 in PBS (3 × 10 min), and blocked for 1 h with 5% normal horse serum (PK-7200, Vector Labs, Burlingame, CA) in PBS. Serial sections were incubated overnight at 4 °C in a solution containing the anti-PV or anti-CB or anti-CR with 2% normal horse serum, 0.05% TX-100 in PBS. Sections were then rinsed in PBS, (3 × 10 min), followed by incubation with biotinylated anti-mouse IgG (5 μg/ml, Vector Labs, Burlingame, CA) or with biotinylated anti-rabbit IgG (5 μg/ml, Vector Labs, Burlingame, CA) (details are reported in Table 3) and then with ABC reagent (Vectastain Kit, PK-7200, Vector Labs, Burlingame, CA). Sections were again rinsed in PBS, for 3 × 10 min. Staining was visualized by incubating the sections in diaminobenzidine (sk-4,105, Vector Labs) solution. The negative controls were performed by replacing either the primary antibodies, anti-mouse/rabbit IgG, or the ABC complex with PBS or non-immune serum. Under these conditions, staining was abolished. Besides, positive controls were carried out testing the primaries antibodies on mouse brain sections. In addition, specificity of the antibodies had already been tested in previous studies (RRID code Table 2 and Table 3).

**Image acquisition and processing**

The stained sections were digitalized using a semi-automated system (D-Sight2, Menarini Diagnistics, Italy) at a magnification of 20 times.

**Tractography**

MRI scans were obtained from the University of Verona with a Bruker tomograph (Bruker, Karlsruhe, Germany) equipped with a 4.7 T, 33-cm bore horizontal magnet (Oxford Ltd., Oxford, UK). Images were acquired with a single-coil configuration. A 7.2 cm inner diameter volume birdcage coil was used as transmitter and receiver. Using a 2D rapid acquisition with relaxation enhancement (RARE) sequence, high-resolution T2w structural images were acquired. Parameters were as follows: repetition time (TR) 35736 ms; echo time (TE) 78,1 ms; field of view (FOV) 6,0 × 5,0 cm; matrix size (MTX)) 240 × 200; 0.250 × 0.250 mm resolution, n. slices 160, 0.5 mm thickness; RARE factor 16; number of averages (NEX) 8; and total acquisition time 1 h and 11 min. Diffusion tensor images were acquired with an Echo Planar Imaging (EPI) sequence. Parameters were as follows: TR 20000 ms, TE 24,7 ms, FOV 6,0 × 5,0 cm; MTX 120 × 100; isotropic in-plane resolution of 0,5 mm; slice thickness 1,0 mm; n. slice 80; EPI factor 11; NEX 6; 30 noncollinear directions acquired with a b value of 3000 s/mm2 and 5 b0 images for a total acquisition time of about 12 h 50 min.

A DTI diffusion scheme was used, and a total of 30 diffusion sampling directions were acquired. The b-value was 3000 s/mm2, with a voxel size of 0.5 x 0.5 x 1 mm. We used then DSI Studio (<http://dsi-studio.labsolver.org>) for the tractography analysis. After computing the diffusion tensor, a deterministic fiber tracking algorithm (Yeh et al., PLoS ONE 8(11): e80713) was used. Ending regions were placed at the right Cl with a volume size of 7e+02 mm2, and left Cl with a volume size of 6.5e+02 mm2. A seeding region was placed in the right primary visual cortex with a volume size of 4.1e03 mm2, and the left primary visual cortex with a volume size of 4.2e03 mm2. The anisotropy threshold was 0.05896. The angular threshold was 45 degrees. The step size was 0.1 mm. The fiber trajectories were smoothed by averaging the propagation direction with 50% of the previous direction. Tracks with length shorter than 2 or longer than 200 mm were discarded. A total of 50000 seeds were placed.

Identification of the presented tracts was achieved by segmenting by hand the visual cortical region, chosen as origin, and the Cl area as the end region, and calculating the white matter tracts running between them. Anatomical references were found in historical papers and recent atlases (Clarke and Whitteridge, 1976; Vanderwolf and Cooley, 2002; Nitzsche et al., 2015)

**Table 2. Primary Antibodies**

|  |  |  |  |
| --- | --- | --- | --- |
| **Antibody** | **Immunogen** | **Manufacturing details** | **Dilution** |
| Anti-PV | PARV-19 hybridoma produced by the fusion of mouse myeloma cells and splenocytes from animmunized mouse. Purified frog muscle parvalbumin was used as the immunogen | Sigma-Aldrich, mouse monoclonal, Clone PARV-19, Product No. P 3088RRID: AB\_477329 | 1:2000 |
| Anti-PV | Produced by hybridization of mouse myeloma cells with spleen cells from mice immunized with parvalbumin purified from carp muscles | Swant, mouse monoclonal,Code No: 235, Lot no: 10-11 (F)RRID: AB\_10000343 | 1:2000 |
| Anti-CR  | The antibody against calretinin was produced in mice by immunization with recombinant human calretinin-22k (identical with calretinin up to Arg178N-terminal). | Swant, mouse monoclonal, Cat# 6B3, Lot n° 010399RRID: AB\_10000320 | 1:1000 |
| Anti-CR | Full length protein | Abcam, rabbit polyclonal, ab702RRID: AB\_305702  | 1:100 |
| Anti-CB | Derived from the CB-955 hybridoma produced by the fusion of mouse myeloma cells and splenocytes from BALB/c mice immunized with a purified bovine kidney calbindin-D-28K | Sigma-Aldrich, mouse monoclonal, Clone CB-955, C9848RRID: AB\_476894 | 1:2000 |
| Anti-CB | This antiserum was produced against recombinant rat calbindin D-28k (CB) | Swant, rabbit polyclonal, Lot No.: 9.03, Code No.: CB-38a RRID: AB\_10000340 | 1:1000 |

**Table 3. Secondary Antibodies**

|  |  |  |  |
| --- | --- | --- | --- |
| **Antibody** | **Type** | **Manufacturing details** | **Dilution** |
| Biotinylated | Anti-mouse IgG (H+L) | Vector Labs, Burlingame, horse,Cat.n. BA-2001, Lot.n. ZC1230RRID: AB\_2336180 | 5 µg/ml |
| Biotinylated | Anti-rabbit IgG (H+L) | Vector Labs, Burlingame, horse,Cat.n. BA-1100, Lot.n. ZA0319RRID: AB\_2336201 | 5 µg/ml |

**RESULTS**

**Claustrum overview**

The rod-like shaped Cl was identified in the ventrolateral telencephalon in the series of rostral coronal blocks, MRI images and in cresyl-violet stained sections (fig. 1 A1-A3). The nucleus then assumed a triangular aspect in the middle region (fig. 1 B1-B3, C1-C3, fig. 2). In the caudalmost region, dorsolateral to the amygdala, the Cl was characterized by a larger and irregular area (fig. 1 D1-D3). Cl was encased in the white matter of the external (Ex) and extreme (Ext) capsules.

**Immunohistochemistry**

Two different antibodies for each CBP were employed and each pair, with the exception of CB, gave the same results. Monoclonal antibody against CB gave negative results both in the Cl and in the visual areas, but clearly stained the Purkinje neurons of the sheep cerebellum (see fig.1 supplementary material). Differently, polyclonal anti-CB immunoreaction was observed both in visual cortex and in Cl.

*Claustrum*

Immunoperoxidase staining in the sheep Cl revealed the presence of both cell bodies and fibers positive to PV, CR and CB (fig. 3). CR immunoreactivity was evenly distributed throughout the dorso-ventral and rostro-caudal extent of the CL while PV and CB stained somata increased moving towards the caudal region. The semi-quantitative representation of the PV, CR, and CB immunostaining distribution is reported in Table 4.

Positive fibers were seen running in all directions throughout the rostro-caudal and the dorso-ventral extent of the CL with a homogeneous distribution. In particular, PV and CB immunoreactive (-ir) fibers were scarce (fig. 3 a-d), while CR-positive fibers made up a dense network surrounding negative cell bodies (fig. 3 e).

|  |  |  |  |
| --- | --- | --- | --- |
| Levels | PV | CR | CB |
| A | - | + | + |
| B | ++ | + | ++ |
| C | +++ | + | +++ |
| D | ++++ | + | +++ |

 **Table 4**. Semi-quantitative representation of PV, CR and CB-ir somata throughout the rostro-caudal levels (A most rostral, D most caudal) of the Cl. -, 0 somata; +, 1-20 somata; ++, 21-40 somata; +++, 41-60 somata; ++++, 61-80 somata.

*Visual area*

The visual area of the ovine cortex was identified based on the available stereotaxic atlases of the species (Richard, 1967; Vanderwolf & Cooley, 2002; Nitzsche et al, 2015) and relevant literature (Rose, 1942; Clarke and Whitteridge, 1976; Clarke et al., 1976; Karamanlidis et al., 1979). The immunocytochemistry of the visual area in the sheep caudal cortex around the lateral sulcus identified largely different GABA-ergic interneuron populations within the cortical thickness (fig. 4).

Calbindin-ir was distributed mostly among layer 3 to 6 (fig. 4E) with a diffuse band of fibers across the layer 3-5 border. Larger somata were present in layer 5 and bipolar bitufted cells were seen in layer 3. Some pyramidal cells were positive to CB immunostaining (fig. 4F). Monoclonal anti-CB did not provide satisfactory staining.

Calretinin-ir neurons were scarce in the V1 area, in which they were found in layer 3 with some also present in layer 5. An extensive neuropil web covered all of the cortical thickness with predominant bands in upper layer 1 and mid-layer 5 (fig. 4H, I). An even clearer fiber band pattern was present in the adjacent peristriate area.

Parvalbumin was very clearly marking layer 3 to 6 with large round somata with vertically oriented dendrites. There were seemingly slightly more cell bodies within layer 5 (fig. 4K). A broad band corresponding to the layer 3-5 border was a major feature of V1. The peristriate area showed no staining (fig. 4C, J, K). Weak positivity could be seen on some pyramidal cells, that could be attributed to dendritic synapsing with PV-ir cells covering the pyramidal somata, or to a markedly lower amount of PV (fig. 4L).

**Tractography**

The tractography algorithm computed between the Cl area and the wider visual cortical area elicited robust fiber tracts following an ipsilateral course (fig. 5). Tracts running directly to the Cl emerged from the occipital end of the visual cortical sensory area (fig. 5A) through a quite homogeneous bundle, travelling along the optic radiation and going through the external capsule region, to reach the claustral territory. Few fibers reached further in the frontal part of the sheep cortex. Despite a large V1 delimitation, only a dorso-lateral, occipital band of fibers were found to reach for the Cl. Seeding from the Cl caused the appearance of additional connection tracts between the caudal part of the Cl to the rostral end of the visual area, towards the rostral end of the marginal sulcus (fig. 5). Although tract endings are not the *forte* of tractography, the tracts reaching the Cl seemed not to end at the dorso-caudal entry from the external capsule, but continue further into the anterior claustral area. We could not find any reliable commissural innervation from V1 to the Cl, while V1-V1 commissural fibers could be detected (not shown).

**DISCUSSION**

In the present study, we have analyzed different neuronal populations in both the Cl and the visual cortex of the sheep, based on the patterns of CBPs expression. We also defined the fiber tracts running between the left and right Cl and the visual areas by MRI scans.

*Choice of markers and methodology*

The choice of CBPs as neurochemical markers of the Cl and visual cortex is based on former studies that indicated that they represent good markers for both structures (for general references see Glezer et al., 1993; Hinova-Palova et al., 2007; Mathur et al., 2009; Mathur, 2014). In particular, PV allows to distinguish the Cl from the endopiriform nucleus (En) and, in fact, in our samples, it was impossible to identify the En in sections immunostained for PV. However, we cannot exclude its presence considering that it has been described in different species (marmoset: Watakabe, 2017; rat: Watson and Puelles, 2017; bat: Orman et al., 2017).

Our experimental series yielded also some unexpected methodological results. The monoclonal antibody anti-CB did not reveal immunoreactivity in the investigated structures, despite having been employed in countless publications (RRID: AB\_476894). Here we emphasize that the same antibody elicited a clear positivity in our positive control, represented by cerebellar Purkinje cells (see supplementary material). One possible explanation might be that there are two different forms of calbindin in the sheep, with a differential expression in the neocortex and the cerebellum, both marked by our polyclonal antibody. And indeed, it is worth noting that in *Ovis aries,* two calbindin CALB1 and CALB2, of 262 aa and 271 aa respectively, are described (UniProt see Ref.). Further investigations are needed to shed light on this scenario, in particular ascribing to the precise antibody target.

*Claustrum*

Our samples showed the unusual shape of the sheep Cl. Indeed, it did not resemble any scheme of Cl type described by Kowianski et al. (1999); however, the shape we observed in the rostral and medial sections was similar to that described in the pig (Pirone et al., 2019). Moreover, the triangular aspect of the middle sheep Cl was a feature found in zebras, lamas and zebus (Buchanan and Johnson, 2011), and also in cats even if the dorsal triangular cat Cl extends ventrally with a thin stem (Sherk, 1986; Johnson et al., 2014).

In our sections and in the 3D model (fig. 2), the sheep Cl appeared as a continuous structure, in contrast from what was reported for the Cl in the gorilla, cetaceans and pigs, where cell islands or lobulations were described (Baizer et al., 2014; Johnson et al., 2014; Pirone et al., 2020). As in other mammal species, the sheep Cl was encased in an external and extreme capsule, the latter lacking or being poorly developed in *Insectivora* and in some rodents (Kowianski et al., 1999); these two capsules make the Cl easily distinguishable from its surrounding structures.

Immunoperoxidase investigation revealed a particular distribution pattern of PV, CR and CB within the sheep Cl. The semi-quantitative representation of CBPs immunoreactive somata showed that PV and CB positive cell bodies increased, moving caudally. Our data show that CR-labeled somata were few and evenly distributed along the rostro-caudal axis, contrarily to what reported for marine Cetartiodactyls, were CR-containing neurons were prevalent in the Cl (Cozzi et al., 2014). Furthermore, PV and CB-ir fibers were scarce while CR-positive fibers formed a dense network surrounding negative cell bodies.

CBPs immunoreactivity was seen throughout the Cl of other mammal species (rat: Druga et al., 1993; monkey: Reynhout & Baizer, 1999; cat: Hinova-Palova et al., 2007; Rahman and Baizer, 2007; human: Hinova-Palova et al., 2014; human, chimpanzee, macaque: Pirone et al., 2014; dog: Pirone et al., 2015; squirrel monkey: Baizer et al., 2020), suggesting that similar subclasses of Cl neurons exist in multiple species and share a homogeneous spatial distribution pattern throughout the anterior-posterior extent of the Cl.

The distribution of CBPs immunoreactivity is not homogeneous along the dorsal-ventral axis of the Cl. In the human Cl, there are more PV-ir elements in the central portion of the nucleus than in the dorsal and ventral aspects (Hinova-Palova et al., 2014). In the cat and dog, the superior Cl contains a higher number of PV-ir somata, and the quantity progressively decreases in the intermediate and inferior parts (Hinova-Palova et al., 2007; Pirone at al., 2015). In human, chimpanzee and macaque Cl both CR- and PV-ir neurons are mostly localized in the central and ventral region of the structure (Pirone et al., 2014). Dorso-ventral subdivisions of the Cl are important, as the dorsal Cl is the part connected to the visual cortex in the cat (LeVay & Sherk, 1981; Minciacchi et al., 1995), but not in primates, where the visual Cl is in the ventral Cl (Remedios et al., 2010).

Our present findings on the distribution of PV-ir cell bodies in the sheep are similar to those we have recently observed in the pig (Pirone at al., 2019), another Cetartiodactyl. In both species, positive somata increased along the rostro-caudal axis of the nucleus. This pattern, considered together with that of CB, might suggest a potentially specific function of the caudal end of the sheep Cl, as already hypothesized for the pig.

*Visual cortex*

The localization of CBPs has been reported in the visual cortex of several mammals, including primates and rodents (for comparative studies see Glezer et al., 1993), carnivores (Yu et al, 2011), bat (Kim et al., 2016) and cetaceans (Hof et al., 1999). However, the presence and distribution of CBPs in the visual cortex of hoofed mammals is seldom reported (Hof et al., 1999), and reflects the reduction of the isocortex to five layers (for general discussion see Cozzi et al., 2017; for the sheep see Peruffo et al., 2019). In our experimental series we found many multipolar and fusiform CB-ir neurons diffuse through the lower layers of the cortex, but not in layer 1, 2 and the upper part of layer 3. The low density of intensely stained CR-ir neurons in our preparations does not reflect the accepted view that the number of CR-positive neurons is much higher than that of CB-positive neurons (Hof et al., 1999). The CR-ir neurons present in layer 2,3 and 5 were however relatively large and their pattern was comparable to what was found in other hoofed mammals as the camel (Hof et al., 1999).

Interestingly, we noted that in the sheep the staining pattern for PV was the opposite of that for CR, which showed a marked stain in the medially adjacent gyrus (cingulate, see fig. 4). Our data present some difference with what described on the distribution and relative presence of CPBs in Cetardiodactyls by Hof et al. in their seminal paper (Hof et al., 1999), to which we refer for comparison. A weak staining of pyramidal cells for CB in layer 3 has been reported previously (Hof et al., 1999) and here we report that PV staining in pyramidal layer 3 is weak too (fig. 4). This points out to the relative similarity in PV and CB distribution, characterized by larger round and multipolar cells for PV, and more bitufted neurons for CB (fig. 4). Interestingly, the staining pattern for PV was the opposite of that of CR, which showed a marked stain in the medially adjacent gyrus (cingulate, see fig. 4).

Neurons expressing CBPs are classically considered GABA-ergic interneurons mainly localized in the neocortex (see Tremblay et al., 2016 for review). Long-range GABA-ergic neurons have been shown to project in cortico-cortical bundles, and in bidirectional hippocampo-entorhinal reciprocal inhibition (Tomioka et al., 2007; Melzer et al., 2012). However, CBPs have also been spotted in projections outside the prosencephalon. PV and CB have been reported in the *substantia nigra*, trigeminal nucleus, and in thalamic neurons that project to the cortex (Gerfen et al., 1985; Bennett-Clarke et al., 1992; Rausell et al., 1992), whilst CR has been found to be dynamically expressed in L5a excitatory pyramidal neurons of the mouse barrel cortex (Liu et al., 2014). Recently, PV-positive excitatory neurons have been identified in the superior colliculus of the mouse, their projections making up the visual pathway to trigger fear (Shang et al., 2019). Moreover, projection neurons of the inferior olive express both CR and CB (Baizer et al., 2011) and the Purkinje cells of the cerebellar cortex express PV (Celio, 1990). Substantiated connections seem to exist in primates (Tanné-Gariépy et al., 2002; Fernàndez-Miranda et al., 2008), cats (LeVay and Sherk, 1981), rabbits (Carman et al., 1964; Gutierrez-Ibarluzea et al., 1999), and sheep, and the Cl itself activates during visuo-motor tasks and integration (Baugh et al. 2011). However, spineless neurons in both in the claustrum and the visual cortex of the cat have been shown to be non-projecting neurons (LeVay and Sherk, 1981). In rabbits, retrograde tracing seemed to demonstrate direct connections from Brodmann’s area 17 by pyramidal cells and stellate cells (Gutierrez-Ibarluzea., 1999). Recent research reports that some claustral cells co-express VGLUT1 and GAD65, suggesting that projecting neurons in the claustrum could provide inhibition to neurons either locally or in more distant cortical areas (Atlan et al., 2018). Although our data does not shine light on any directionality, as a working hypothesis, we may consider that part of that input to the Cl in the sheep could be inhibitory and mediated by GABA-ergic neurons.

*Tractography*

Tractography has been applied to map human cortico-claustral connections and reported the presence of occipito-frontal fascicles coming from the occipito-parietal region to the Cl and beyond (Fernàndez-Miranda et al., 2008; Meola et al., 2015; Fernandez-Miranda et al., 2015). This is in agreement with our more focused DTI which was limited to the primary visual area in the dorso-occipital region of the sheep brain and its connection to the Cl. The emerging bundle from V1 seems to concord with Clarke and Whitterige’s (1976) reduced V1 findings. Our data therefore confirm the existence of such a bundle seems in the sheep. As was shown in the mouse (Atlan et al., 2017), our tracts reveal a rather central, tight bundle reaching the claustrum (Figure 5). However, we detected no contralateral projection in the visuo-claustral interconnections. In all of our subjects used for tractography (n = 3), a slight inconstant asymmetry was observed, which could have resulted from the variability in hand parsing of the cortical areas, or specimen variability, but no anatomical feature should be deducted from it. Given the fact that the regions were segmented by hand, there is a risk to catch unrelated passing fibers, which is especially problematic in the white-matter embedded claustrum. However, this is applicable to any similar study, and we did our best to be conservative in the segmentation. A known limitation of tractography it the estimation of tract endings, and most importantly, directionality. There seems to be evidence, however, that a visual to claustrum connection exists, at least in the mouse (Wang et al., 2016). Finally, in the cat, the visual zone is sitting at the dorsocaudal part of the clausrtum (Sherk, 1986), which is coherent with the entry level of our tracts (Figure 5E).

Sheep and goats have laterally placed eyes and a limited binocular vision, but they do possess binocular neurons (Clarke and Whitteridge, 1976; Clarke et al., 1976) and are known to have a good depth perception (Walk and Gibson, 1961). Contralateral claustro-visual projections have long been described in the cat (Jayaraman and Updyke, 1979; Sanides and Buchholtz, 1979; Olson and Graybiel, 1980; Squatrito et al., 1980). We lowered the thresholds used for the ipsilateral bundles in the attempt to find crossing fibers reaching the contralateral Cl, but with no result. Our data suggest that contralateral connections are absent in the sheep, or perhaps very limited as in the cat (Olson & Graybiel, 1980). We also note that connections (seeding from the claustrum, fig. 5F), may reach peristriate areas such as V2, V3 or V4 that we did not map, because their precise location and topography has not been identified in the sheep, although tracing studies showed a quite wide visual territory (Karamanlidis et al., 1979). The absence of contralateral projections could be of prime importance if confirmed by other studies, to rule out the role of the Cl in stereopsis.

**REFERENCES**

Atlan, G., Terem, A., Peretz-Rivlin, N., Groysman, M., and Citri, A. (2017). Mapping synaptic cortico-claustral connectivity in the mouse. *J. Comp. Neurol.* 525, 1381–1402. doi:10.1002/cne.23997.

Atlan, G., Terem, A., Peretz-Rivlin, N., Sehrawat, K., Gonzales, B. J., Pozner, G., Tasaka, G. ichi, Goll, Y., Refaeli, R., Zviran, O., Lim, B. K., Groysman, M., Goshen, I., Mizrahi, A., Nelken, I., & Citri, A. (2018). The Claustrum Supports Resilience to Distraction. *Current Biology*, *28*(17), 2752-2762.e7. <https://doi.org/10.1016/j.cub.2018.06.068>

Baizer JS. 2001. Serotonergic innervation of the primate claustrum. Brain Res Bull 55:431-434

Baizer, J.S., Webster, C.J., Baker, J.F., 2020. The Claustrum in the Squirrel Monkey. Anat Rec 303:1439–1454.

Baizer, J.S., Sherwood, C.C,, Hof, P.R., Witelson, S.F., Sultan, F., 2011. Neurochemical and structural organization of the principal nucleus of the inferior olive in the human. Anat Rec (Hoboken) 294:1198-1216.

Baizer JS, Sherwood CC, Noonan M, Hof PR. Comparative organization of the claustrum: what does structure tell us about function? Front Syst Neurosci. 2014 Jul 2;8:117. doi: 0.3389/fnsys.2014.00117.

Baugh, L. A., Lawrence, J. M., & Marotta, J. J. (2011). Novel claustrum activation observed during a visuomotor adaptation task using a viewing window paradigm. *Behavioural brain research*, *223*(2), 395-402.

Bennett-Clarke CA, Chiaia NL, Jacquin MF, Rhoades RW. 1992. Parvalbumin and calbindin immunocytochemistry reveal functionally distinct cell groups and vibrissa-related patterns in the trigeminal brainstem complex of the adult rat. J Comp Neurol 320:323–338.

Binks, D., Watson, C., Puelles, L., 2019. A Re-evaluation of the Anatomy of the Claustrum in Rodents and Primates—Analyzing the Effect of Pallial expansion. Frontiers in Neuroanatomy, 13, 34

Buchanan, K.J., Johnson, J.I., 2011. Diversity of spatial relationships of the claustrum and insula in branches of the mammalian radiation. Ann. N.Y. Acad. Sci. 1225(Supp. 1), E30–E63. doi: 10.1111/j.1749-6632.2011. 06022.x

Butler, A.B., Reiner, A., Karten, H.J., 2011. Evolution of the amniote pallium and the origins of mammalian neocortex. Ann. N.Y. Acad. Sci. 1225, 14–27. http://dx.doi.org/10.1111/j.1749-6632.2011.06006.x.

Carey RG, Neal TL. 1986. Reciprocal connections between the claustrum and visual thalamus in the tree shrew (*Tupaia glis*). Brain Res 386:155-168.

Carman, J.B., Cowan, W.M., Powell, T.P., 1964. the Cortical Projection Upon the Claustrum. J. Neurol. Neurosurg. Psychiatry 27, 46–51. https://doi.org/10.1136/jnnp.27.1.46

Celio MR. 1990. Calbindin D-28k and parvalbumin in the rat nervous system. Neuroscience 35:375-475.

Clarke, P.G.H., Whitteridge, D., 1976. The cortical visual areas of the sheep. J. Physiol. 256, 497–508. https://doi.org/10.1113/jphysiol.1976.sp011335

Clarke,P.G.H., Donaidson,I.M.L., Whitteridge,D., 1976. Binocular visual mechanisms in cortical areas I and II of the sheep. J. Physiol. (Lond.) 256, 509-526.

Cozzi, B., Roncon, G., Granato, A., Giurisato, M., Castagna, M., Peruffo, A., Panin, M., Ballarin, C., Montelli, S., Pirone, A., 2014. The claustrum of the bottlenose dolphin Tursiops truncatus (Montagu 1821). Front. Syst. Neurosci. 8, 42. <https://doi.org/10.3389/fnsys.2014.00042>

Cozzi B, De Giorgio A, Peruffo A, Montelli S, Panin M, Bombardi C, Grandis A, Pirone A, Zambenedetti P, Corain L, Granato A. 2017. The laminar organization of the motor cortex in monodactylous mammals: a comparative assessment based on horse, chimpanzee, and macaque. Brain Struct Funct 222: 2743–2757. https://doi.org/10.1007/s00429-017-1397-z

Crick, F.C., Koch, C., 2005. What is the function of the claustrum? Philos. Trans. R. Soc. B Biol. Sci. 360, 1271–1279. https://doi.org/10.1098/rstb.2005.1661

Day-Brown JD, Slusarczyk AS, Zhou N, Quiggins R, Petry HM, Bickford ME. 2016. Synaptic organization of striate cortex projections in the tree shrew: a comparison of the claustrum and dorsal thalamus. J Comp Neurol 52:1403–1420

Deutch AY, Mathur BN (2015) Editorial: the CL: charting a way forward for the brain’s most mysterious nucleus. Front Syst Neurosci 9, 103.

Druga R, Chen S, Bentivoglio M. 1993. Parvalbumin and calbindin in the rat claustrum: an immunocytochemical study combined with retrograde tracing frontoparietal cortex. J Chem Neuroanat 6: 399–406.

Edelstein LR, Denaro FJ. 2004. The claustrum: a historical review of its anatomy, physiology, cytochemistry and functional significance. Cell Mol Biol (Noisy-le-grand) 50:675-702.

Eiden LE, Mezey E, Eskay RL, Beinfeld MC, Palkovits M. 1990. Neuropeptide content and connectivity of the rat claustrum. Brain Res 523:245-250.

Fernández-Miranda, J.C., Rhoton, A.L., Álvarez-Linera, J., Kakizawa, Y., Choi, C., De Oliveira, E.P., 2008. Three-dimensional microsurgical and tractographic anatomy of the white matter of the human brain. Neurosurgery 62, 989–1028. https://doi.org/10.1227/01.NEU.0000297076.98175.67

Fernández-Miranda, J.C., Rhoton, A.L., Kakizawa, Y., Choi, C., Álvarez-Linera, J., 2008. The claustrum and its projection system in the human brain: A microsurgical and tractographic anatomical study - Laboratory investigation. J. Neurosurg. 108, 764–774. https://doi.org/10.3171/JNS/2008/108/4/0764

Fernández-Miranda, J.C., Wang, Y., Pathak, S., Stefaneau, L., Verstynen, T., Yeh, F.C., 2015. Asymmetry, connectivity, and segmentation of the arcuate fascicle in the human brain. Brain Struct. Funct. 220, 1665–1680. https://doi.org/10.1007/s00429-014-0751-7

Gerfen CR, Baimbridget KG, Miller JJ. 1985. The neostriatal mosaic: Compartmental distribution of calcium-binding protein and parvalbumin in the basalganglia of the rat and monkey. Proc Natl Acad Sci U S A 82:8780–8784.

Glezer, I.I., Hof, P.R., Leranth, C., Morgane, P.J., 1993. Calcium- binding protein-containing neuronal populations in mammalian visual cortex: A comparative study in whales, insectivores, bats, rodents, and primates. Cereb. Cortex 3, 249–272

Goll, Y., Atlan, G., Citri, A., 2015. Attention: The claustrum. Trends Neurosci. 38, 486–495. <https://doi.org/10.1016/j.tins.2015.05.006>

Gutierrez-Ibarluzea I., Acera-Osa A., Mendizabal-Zubiaga J.L., Arana-Arri E., Bueno-Lopez J.L., Reblet C. Eur J Anat, 3 (2): 101-109 (1999). Morphology and laminar distribution of cortico-claustral neurons in different areas of the rabbit cerebral cortex.

Hinova-Palova DV, Edelstein LR, Paloff AM, Hristov S, Papantchev VG, Ovtscharoff WA. 2007 Parvalbumin in the cat claustrum: Ultrastructure, distribution and functional implications. Acta Histochem 109: 61—77

Hinova-Palova D, Edelstein L, Papantchev V, Landzhov B, Malinova L, Todorova-Papantcheva D, Minkov M, Paloff A, Ovtscharoff W. 2012. Light and electron-microscopic study of leucine enkephalin immunoreactivity in the cat claustrum. Journal of molecular histology 43:641-649.

Hinova-Palova D, Edelstein L, Paloff A, Hristov S, Papantchev V, Ovtscharoff W. 2008. Neuronal nitric oxide synthase immunopositive neurons in cat claustrum--a light and electron microscopic study. Journal of molecular histology 39:447-457.

Hinova-Palova DV, Edelstein L, Landzhov B, Minkov M, Malinova L, Hristov S, Denaro FJ, Alexandrov A, Kiriakova T, Brainova I, Paloff A, Ovtscharoff W. Topographical distribution and morphology of NADPH-diaphorase-stained neurons in the human claustrum. Front Syst Neurosci. 2014a May 27;8:96. doi: 10.3389/fnsys.2014.00096.

Hinova-Palova DV, Edelstein L, Landzhov BV, Braak E, Malinova LG, Minkov M, Paloff A, Ovtscharoff W. 2014b. Parvalbumin immunoreactive neurons in the human CL. Brain Struct Funct 219:1813–1830.

Hinova-Palova D, Kotov G, Landzhov B, Edelstein L, Iliev A, Stanchev S, Georgiev GP, Kirkov V, Angelov T, Nikolov D, Fakih K, Paloff A. Cytoarchitecture of the dorsal claustrum of the cat: a quantitative Golgi study. J Mol Histol. 2019a Oct;50(5):435-457. doi: 10.1007/s10735-019-09839-7.

Hinova-Palova D, Landzhov B, Iliev A, Kotov G, Stanchev S, Kirkov V, Georgiev GP, Edelstein L, Paloff A. Ultrastructure of the dorsal claustrum in cat. II. Synaptic organization. Acta Histochem. 2019b May;121(4):383-391. doi: 10.1016/j.acthis.2019.02.009.

Hof PR, Glezer II, Condé F, Flagg RA, Rubin MB, Nimchinsky EA, Vogt Weisenhorn DM (1999) Cellular distribution of the calcium-binding proteins parvalbumin, calbindin, and calretinin in the neocortex of mammals: phylogenetic and developmental patterns. J Chem Neuroanat 16: 77-116.

Jackson,J., Smith, J.B., Lee, K.B., 2020. The Anatomy and Physiology of Claustrum-Cortex Interactions. *Annual Review of Neuroscience,* 43:231–47.

Jayaraman, A., Updyke, B. V, 1979. Organization of visual cortical projections to the claustrum in the cat. Brain Res. 178, 107–115. [https://doi.org/10.1016/0006-8993(79)90091-X](https://doi.org/10.1016/0006-8993%2879%2990091-X)

Karamanlidis, A. N., Saigal, R. P., Giolli, R. A., Mangana, O., & Michaloudi, H. (1979). Visual thalamocortical connections in sheep studied by means of the retrograde transport of Horseradish‐Peroxidase. *Journal of Comparative Neurology*, *187*(2), 245–259. https://doi.org/10.1002/cne.901870202

Kim, H.-G., Gu, Y.-N., Lee, K.-P., Lee, J.-G., Kim, C.-W., Lee, J.-W., et al. (2016). Immunocytochemical localization of the calcium-binding proteins calbindin D28K, calretinin, and parvalbumin in bat visual cortex. *Histology and histopathology*, *31*(3), 317–27. <https://doi.org/10.14670/HH-11-680>

Kowianski P, Dziewiatkowski J, Kowianska J, Morys J. 1999. Comparative anatomy of the claustrum in selected species: A morphometric analysis. Brain Behav Evol 53:44–54.

Kowiański P, Morys JM, Dziewiatkowski J, Wojcik S, Sidor-Kaczmarek J, Morys J. 2008. NPY- , SOM- and VIP-containing interneurons in postnatal development of the rat claustrum. Brain Res Bull 76:565-571.

Krimmel SR, Qadir H, Hesselgrave N, White MG, Reser DH, Mathur BN, Seminowicz DA. 2019. Resting state functional connectivity of the rat claustrum. Front. Neuroanat. 13:22. doi: 10.3389/fnana.2019.00022

Landzhov B, Hinova-Palova D, Edelstein L, Dzhambazova E, Brainova I, Georgiev GP, Ivanova V, Paloff A, Ovtscharoff W. 2017. Comparative investigation of neuronal nitric oxide synthase immunoreactivity in rat and human claustrum. J Chem Neuroanat 86:1-14.

LeVay, S., Sherk, H., 1981. The visual claustrum of the cat. I. Structure and connections. J. Neurosci. 1, 956–980. https://doi.org/10.1523/jneurosci.01-09-00956.1981

Lipowska M, Kowianski P, Majak K, Jagalska-Majewska H, Morys J. 2000. The connections of the endopiriform nucleus with the insular claustrum in the rat and rabbit. Folia Morphol (Warsz) 59:77-83.

Liu, J., Liu, B., Zhang, X.Y., Yu, B., Guan, W., Wang, K., Yang, Y., Gong, Y., Wu, X., Yanagawa, Y., Wu, S., Zhao, C., 2014. Development of the paralemniscal pathway in the barrel cortex. Mol. Brain 7:1–13. doi: 10.1186/s13041-014-0084-8

Maçarico da Costa N, Fürsinger D, Martin KAC. 2010. The synaptic organization of the claustral projection to the cat’s visual cortex. J. Neurosci. 30: 13166 –13170.

Mathur, B.N., 2014. The claustrum in review. Front. Syst. Neurosci. 8, 1–11. <https://doi.org/10.3389/fnsys.2014.00048>

Mathur BN, Caprioli RM, Deutch AY. 2009. Proteomic analysis illuminates a novel structural definition of the claustrum and insula. Cereb Cortex 19:2372–2379.

Melzer, S. et al. 2012. Long-range-projecting GABAergic neurons modulate inhibition in hippocampus and entorhinal cortex. Science 335, 1506–1510

Meola, A., Comert, A., Yeh, F.C., Stefaneanu, L., Fernandez-Miranda, J.C., 2015. The controversial existence of the human superior fronto-occipital fasciculus: Connectome-based tractographic study with microdissection validation. Hum. Brain Mapp. 36, 4964–4971. <https://doi.org/10.1002/hbm.22990>

Minciacchi D, Granato A, Antonini A, Tassinari G, Santarelli M, Zanolli L, Macchi G. 1995. Mapping subcortical extrarelay afferents on to primary somatosensory and visual areas in cats. J Comp Neurol362: 46–70. doi:10.1002/cne. 903620104

Narikiyo, K., Mizuguchi, R., Ajima, A., Shiozaki, M., Hamanaka, H., Johansen, J. P., Mori, K., & Yoshihara, Y. (2020). The claustrum coordinates cortical slow-wave activity. *Nature Neuroscience*, *23*(June). <https://doi.org/10.1038/s41593-020-0625-7>

Nitzsche, B., Frey, S., Collins, L.D., Seeger, J., Lobsien, D., Dreyer, A., Kirsten, H., Stoffel, M.H., Fonov, V.S., Boltze, J., 2015. A stereotaxic, population-averaged T1w ovine brain atlas including cerebral morphology and tissue volumes. Front. Neuroanat. 9, 1–14. https://doi.org/10.3389/fnana.2015.00069

Norita, M. (1977). Demonstration of bilateral claustro-cortical connections in the cat with the method of retrograde axonal transport of horseradish peroxidase. *Archivum histologicum japonicum*, *40*(1), 1-10.

Olson, C. R., & Graybiel, A. M. (1980). Sensory maps in the claustrum of the cat. *Nature*, *288*(5790), 479-481.

Orman R, Kollmar R, Stewart M. 2017. Claustrum of the short-tailed fruit bat, Carollia perspicillata: alignment of cellular orientation and functional connectivity. J Comp Neurol 525:1459–1474.

Peruffo, A., Corain, L., Bombardi, C., Centelleghe, C., Grisan, E., Graïc, J.-M., Bontempi, P., Grandis, A., Cozzi, B., 2019. The motor cortex of the sheep: laminar organization, projections and diffusion tensor imaging of the intracranial pyramidal and extrapyramidal tracts. Brain Struct. Funct. 224, 1933–1946. https://doi.org/10.1007/s00429-019-01885-x

Pirone, A., Castagna, M., Granato, A., Peruffo, A., Quilici, F., Cavicchioli, L., Piano, I., Lenzi, C., Cozzi, B., 2014. Expression of calcium-binding proteins and selected neuropeptides in the human, chimpanzee, and crab-eating macaque claustrum. Front. Syst. Neurosci. 8, 191–192. <https://doi.org/10.3389/fnsys.2014.00099>

Pirone A, Magliaro C, Giannessi E, Ahluwalia A. 2015. Parvalbumin expression in the claustrum of the adult dog. An immunohistochemical and topographical study with comparative notes on the structure of the nucleus. J Chem Neuroanat 64–65:33–42.

Pirone A, Cozzi B, Edelstein L, et al. (2012) Topography of Gng2- and NetrinG2-expression suggests an insular origin of the human CL. PLoS ONE 7(9), e44745.

Pirone, A., Miragliotta, V., Ciregia, F., Giannessi, E., Cozzi, B., 2018. The catecholaminergic innervation of the claustrum of the pig. J. Anat. 232, 158–166. https://doi.org/10.1111/joa.12706

Pirone, A., Miragliotta, V., Cozzi, B., Granato, A., 2019. The Claustrum of the Pig: An Immunohistochemical and a Quantitative Golgi Study. Anat. Rec. 302, 1638–1646. <https://doi.org/10.1002/ar.24073>

Pirone, A., Lazzarini, G., Lenzi, C., Giannessi, E., Miragliotta, V., 2020. Immunolocalization of cannabinoid receptor 1 (CB1), monoglyceride lipase (MGL) and fatty-acid amide hydrolase 1 (FAAH) in the pig claustrum. Journal of Chemical Neuroanatomy <https://doi.org/10.1016/j.jchemneu.2020.101843>

Puelles, L., Ayad, A., Sandoval, J.E., Alonso, A., Medina, L., Ferran, J.L., 2016a. Selective early expression of the orphan nuclear receptor Nr4a2 identifies the claustrum homolog in the avian mesopallium; impact on sauropsidian/mammalian pallium comparisons. J. Comp. Neurol. 524, 665–703.

Rahman FE, Baizer JS. 2007. Neurochemically defined cell types in the claustrum of the cat. Brain Res 1159:94–111.

Rausell E, Bae CS, Viñuela A, Huntley GW, Jones EG. 1992. Calbindin and parvalbumin cells in monkey VPL thalamic nucleus: distribution, laminar cortical projections, and relations to spinothalamic terminations. J Neurosci 12:4088–4111.

Reynhout K, Baizer JS. 1999. Immunoreactivity for calciumbinding Proteins in the claustrum of the monkey. Anat Embryol 199:75–83.

Remedios R, Logothetis NK, Kayser C. 2010.Unimodal responses prevail within the multisensory claustrum. *J. Neurosci.* 30, 12902–12907. doi:10. 1523/JNEUROSCI.2937-10.2010

Reser DH, Majka P, Snell S, Chan JM, Watkins K, Worthy K, Quiroga MD, Rosa MG. 2017. Topography of claustrum and insula projections to medial prefrontal and anterior cingulate cortex of the common marmoset (Callithrix jacchus). J Comp Neurol 525:1421–1441.

Richard P (1967) Atlas stéréotaxique du cerveau de Brebis “Préalpesdu-Sud”. Institut National de la Recherche Agronomique, Paris

Riche, D., & Lanoir, J. (1978). Some claustro‐cortical connections in the cat and baboon as studied by retrograde horseradish persocidase transport. *Journal of Comparative Neurology*, *177*(3), 435-444.

Rose, J. E. (1942). A cytoarchitectural study of the sheep cortex. *The Journal of Comparative Neurology*, *76*(1), 1–55. https://doi.org/10.1002/cne.900760102

Sanides D, Buchholtz cs. 1979. Identificationof the projection from the visual cortex to the claustrum by anterograde axonal transport in the cat.Exp Brain Res 34: 197-200.

Shang, C., Liu, Z., Chen, Z., Shi, Y., Wang, Q., Liu, S., Li, D., Cao, P., 2019. A parvalbumin-positive excitatory visual pathway to trigger fear responses in mice. Science, 348:1472-1477.

Sherk, H. (1986). The Claustrum and the Cerebral Cortex. In E. G. Jones & A. Peters (Eds.), *Sensory-Motor Areas and Aspects of Cortical Connectivity. Cerebral Cortex, vol 5.* (pp. 467–499). Springer. <https://doi.org/10.1007/978-1-4613-2149-1_13>

Squatrito S, Battaglini PP, Galletti C, Riva Sanseverino E. 1980. Projections from the visual cortex to the contralateral claustrum of the cat revealed by an anterograde axonal transport method. Neurosci Lett 19: 271-275.

Tanné‐Gariépy, J., Boussaoud, D., & Rouiller, E. M. (2002). Projections of the claustrum to the primary motor, premotor, and prefrontal cortices in the macaque monkey. *Journal of Comparative Neurology*, *454*(2), 140-157.

Tomioka, R. & Rockland, K. S. 2007, Long-distance corticocortical GABAergic neurons in the adult monkey white and gray matter. J. Comp. Neurol. 505, 526–538.

Tremblay, R., Lee, S., Rudy, B., 2016. GABAergic Interneurons in the Neocortex: From Cellular Properties to Circuits. Neuron 91(2): 260–292. doi:10.1016/j.neuron.2016.06.033.

UniProt website (<https://www.uniprot.org/uniprot/?query=calbindin+sheep&sort=score>), accessed on 07/04/2020.

Vanderwolf, C.H., Cooley, R.C., 2002. The Sheep Brain: A Photographic Series, 2nd ed. A J Kirby & Co.

Vertes RP, Hoover WB. 2008. Projections of the paraventricular and paratenial nuclei of the dorsal midline thalamus in the rat. J Comp Neurol 508:212-237

Walk,R.D., Gibson,E.J.: A comparative and analytical study of visual depth perception. Psychol. Monogr.75, 1-44 (1961)

Wang Q, Ng L, Harris JA, Feng D, Li Y, Royall JJ, Oh SW, Bernard A, Sunkin SM, Koch C, et al. 2017. Organization of the connections between claustrum and cortex in the mouse. J Comp Neurol 1346:1317–1346.

Watakabe A. 2017. In situ hybridization analyses of claustrumenriched genes in marmosets. J Comp Neurol 525:1442–1458.

Watson, C., & Puelles, L. (2017). Developmental gene expression in the mouse clarifies the organization of the claustrum and related endopiriform nuclei. *Journal of Comparative Neurology*, *525*(6), 1499–1508. https://doi.org/10.1002/cne.24034

White, M.G., Mathur, B.N., 2018. Frontal cortical control of posterior sensory and association cortices through the claustrum. Brain Struct. Funct. 223, 1–8. <https://doi.org/10.1007/s00429-018-1661-x>

Yeh, F.C, Verstynen, T.D, Wang, Y., Fernández-Miranda, J.C., Tseng, W.Y.I, 2013. Deterministic Diffusion Fiber Tracking Improved by Quantitative Anisotropy. PLoS ONE 8(11): e80713.

Yu, S.-H., Lee, J.-Y., & Jeon, C.-J. (2011). Immunocytochemical Localization of Calcium-Binding Proteins, Calbindin D28K-, Calretinin-, and Parvalbumin-Containing Neurons in the Dog Visual Cortex. *Zoological Science*, *28*(9), 694–702. <https://doi.org/10.2108/zsj.28.694>

**FIGURE LEGENDS**

**Figure 1.** Anterior (A) to posterior (D) coronal aspects of the sheep claustrum. Fresh coronal blocks (first row A1 to D1; scale bar = 5 mm); MRI digital sections (second row, scale bar = 1 cm); Cresyl violet staining (third row; scale bar = 2 mm). Cl (cl) outline is drawn by red lines. a, amygdala; aha, amygdalohippocampal area; c, caudate; cp, caudoputamen; Ex, external capsule; Ext, extreme capsule; p, putamen.

**Figure 2**. 3D reconstruction of the sheep claustrum in red. A, anterior; D, dorsal; L, left; P, posterior; R, right; V, ventral.

**Figure 3.** Immunohistochemical staining of the sheep Cl. Immunoperoxidase reaction shows the distribution of CB in the rostral (A) and caudal (B) parts, PV in the rostral (C) and caudal (D) parts, and CR caudal part (E) immunoreactivity in the Cl (dashed line). Below A, very few and weakly stained CB-ir cells are spotted. Higher magnification of the frames below B display CB-ir cells and scarce fibers (enlarged in the inset). Below C are rare fibers with no real soma stain for PV. Below D, higher magnification shows a moderate density of PV-ir fibers and two positive neurons (enlarged in the inset). Below E, higher magnification shows a dense network of CR-positive fibers (enlarged in the inset). Cl, claustrum; ctx, cortex; Pt, putamen. Scale bars = 500 μm (upper row), 100 μm (lower row), 10 μm (insets).

**Figure 4.** Immunocytochemical staining of the sheep V1. Immunocytochemistry reveals the patterns of calcium binding proteins in V1 (E, H, K) and the peristriate area (D, G, J). Calbindin is found in most of the cortical thickness (A, D, E, F), staining round interneurons and some pyramidal cells, while calretinin is much more localized, notably in layer 1 (B, G, H, I). Parvalbumin shows also a large positivity throughout the cortical thickness (C, J, K, L) with a marked neuropil band in V1 (K). Positive cells are rather large and round multipolar, with weaker staining of some pyramidal neurons (L). Bar in A, B, C is 1000 µm; bar in D, E, G, H, J and K is 500 µm., F, I, L bar is 50 µm.

**Figure 5.** Representation of fiber tracts running between the left and right Cl (yellow and red, respectively) and the left and right visual areas (in blue and green, respectively). MRI images represent the orthogonal planes. A, Left antero-lateral view. Indicative bar = 1 cm. B, Right antero-lateral view. In A and B, the extent of the tracts in the claustra can be appreciated, as well as the posterior access of the fibers, originating from the occipital aspect of the visual cortex. C, Left Lateral view. D, Right postero-lateral view. E, Ventral view where the left tracts can be seen reaching up to the visual cortex in the dorso-occipital cortex. F, Left antero-lateral view, where the Cl is used as a seed to map the tracts originating from it. The fibers exiting the Cl are very rich. Note the projection to the anterior part of the visual territory, emerging from the caudo-dorsal part of the Cl.