***HFE* mRNA expression is responsive to intracellular and extracellular iron loading: short communication**

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**Abstract**

**Background:** In liver hepatocytes, the *HFE* gene regulates cellular and systemic iron homeostasis by modulating cellular iron-uptake and producing the iron-hormone hepcidin in response to systemic iron elevation. However, the mechanism of iron-sensing in hepatocytes remain enigmatic. Therefore, to study the effect of iron on *HFE* and hepcidin *(HAMP*) expressions under distinct extracellular and intracellular iron-loading, we examined the effect of holotransferrin treatment (1,2, 5 and 8 g/L for 6 h) on intracellular iron levels, and mRNA expressions of *HFE* and *HAMP* in wild-type HepG2 and previously characterized iron-loaded recombinant-TfR1 HepG2 cells.

**Methods and Results:** Gene expression was analyzed by real-time PCR and intracellular iron was measured by ferrozine assay. Data showed that in the wild-type cells, where intracellular iron content remained unchanged, *HFE* expression remained unaltered at low holotransferrin treatments but was upregulated upon 5 g/L (p<0.04) and 8 g/L (p=0.05) treatments. *HAMP* expression showed alternating elevations and increased upon 1 g/L (p<0.05) and 5 g/L (p<0.05). However, in the recombinant cells that showed higher intracellular iron levels than wild-type cells, *HFE* and *HAMP* expressions were elevated only at low 1 g/L treatment (p<0.03) and were repressed at 2 g/L (p<0.03) treatment. Under holotransferrin-untreated conditions, the iron-loaded recombinant cells showed higher expressions of *HFE* (p<0.03) and *HAMP* (p=0.05) than wild-type cells.

**Conclusions:** *HFE* mRNA was independently elevated by extracellular and intracellular iron-excess. Thus, it may be involved in sensing both, extracellular and intracellular iron. Repression of *HAMP e*xpression under simultaneous intracellular and extracellular iron-loading resembles certain iron-excess pathologies.

**Keywords**: HFE, iron-sensing, holotransferrin, hepcidin

**Introduction**

Maintenance of cellular and systemic iron homeostasis in the body is a dynamic process involving several signal transduction pathways. The hemochromatosis protein HFE maintains body iron homeostasis by participating in the induction of hepcidin (*HAMP*), the systemic iron regulator, by a yet incompletely understood mechanism [1–3]. Mutations in iron-related genes such as *HFE, HJV*, *HAMP* and *TFR2* cause diminished hepcidin production which results in systemic and tissue iron overload, referred as type 1, type 2A, type 2B and type 3 hereditary hemochromatosis, respectively [4]. However, despite the presence of functional wild-type alleles of these genes, low to moderate tissue iron excess is also observed in alcoholic liver disease, hepatitis C infections, non-alcoholic fatty liver disease, non-alcoholic steatohepatitis and type 2 diabetes [5–8]. In these cases, iron loading can exacerbate the pathophysiology via excess-iron-induced oxidative stress [9]. Thus, it is important to fully delineate the iron-sensing mechanisms to formulate therapeutic interventions, particularly for the low-moderate iron-loaded conditions where, unlike hereditary hemochromatosis, phlebotomy is not practiced for removal of excess iron.

The mRNA response of *HFE* to a range of increasing extracellular iron, elevated intracellular iron and its relationship with *HAMP* expression has not been studied so far. Hence, in this short study, we investigated the effect of a range of holotransferrin (holo-Tf) concentrations (1 to 8 g/L) on *HFE* and *HAMP* mRNA expressions, and intracellular iron content. First, we observed these responses in the wild type (Wt) HepG2 cells, where holo-Tf supplementation represent physiological conditions with extracellular (systemic) iron elevation prior to intracellular/tissue iron loading. Then, we examined the responses in the previously characterized recombinant (rec)-TfR1 HepG2 cells [10]. As these cells can achieve intracellular iron overloading [10], holo-Tf supplementation to these cells represent pathological conditions, which show simultaneously increased extracellular (systemic) and intracellular iron levels. Finally, to understand the exclusive effect of high intracellular iron content, we compared the expression levels between holo-Tf-untreated Wt and recombinant cells. Unlike most previous holo-Tf supplementation studies that were conducted at longer time-points of 24 h, 48 h or 72 h [11–13], here, we studied the effect following 6 h of holotransferrin treatment to examine early responses.

**Materials and Methods**

**Cell culture and treatments**

Maintenance and holo-Tf supplementation to the WtHepG2 cells (Health Protection Agency, UK) and rec-TfR1HepG2 cells was as described previously [10]. Cells were treated with holo-Tf (1, 2, 5 and 8 g/L) prepared in serum-free EMEM (0 g/L) for 6 h and assessed for various parameters. As treatment with 8 g/L holo-Tf represent very high holo-Tf concentrations and the rec-TfR1 HepG2 cells had the potential for intracellular iron-overloading following holo-Tf supplementation [10], the effect of this concentration was studied only in Wt cells.

**Determination of intracellular iron content**

Cellular iron content determined by ferrozine assay [14] was normalized to protein, content, as quantified by Bradford method. Iron levels were expressed as nmoles iron/mg protein.

**Gene expression analysis**

Primers (Invitrogen, UK) for expression analyses, RNA extraction, cDNA conversion and assessment for mRNA expression via real-time PCR by using Quantifast SYBR green kit (Qiagen, UK), was as previously described [10,15].Data was analyzed by the relative quantification method, Delta-Delta Ct (∆∆Ct) and expressed as 2 -∆∆Ct[16].

**Statistical analysis**

Data analysis was performed using one-way ANOVA. The level of significance was set at p<0.05. Data was presented as mean ± SEM (n=3).

**Results**

In the Wt cells, while intracellular iron content remained unaltered following holo-Tf supplementation (Fig.1a), *HFE* mRNA expression significantly increased by 3.5-fold (p<0.04) upon 5 g/L treatment and further increased by 4.5-fold (p=0.05) upon 8 g/L treatment (Fig.1b). Expression levels remained unaltered at lower concentrations of 1 g/L and 2 g/L (Fig.1b). Differentially, *HAMP* expression showed a pattern of alternating responses i.e. a significant 1.8-fold (p<0.05) up-regulation upon 1 g/L treatment, unaltered expression upon 2 g/L treatment followed by a significant 2.3-fold up-regulation upon 5 g/L treatment (p<0.05) and then, down-regulation upon 8 g/L treatment (Fig.1c).

Prior to expression studies in the recombinant cells, intracellular iron loading was confirmed. Data showed that following treatment with most holo-Tf concentrations,

intracellular iron content in these cells was higher than Wt cells (Figs. 2a and 1a).In the recombinant cells, over the increasing holo-Tf concentrations, although intracellular iron content decreased at 2 g/L (p<0.01), it increasedat 5 g/L treatment (p<0.03) that restored the high levels, as in untreated conditions (Fig.2a).These cells differed from the Wt cells in *HFE* and *HAMP* expression patterns (summarized in Table 1). Here, *HFE* expression increased upon 1 g/L (p=0.07), but then decreased upon 2 g/L holo-Tf treatment (p<0.03), and remained unaltered at 5 g/L (Fig.2b). Similarly, *HAM*P expression increased by 3.5-fold at 1 g/L (p<0.03) followed by arepression at 2 g/L (p<0.03) and remained unaltered at 5 g/L treatment (Fig.2c).

Further, to understand the exclusive effect of intracellular iron loading, *HFE* and *HAMP* expressions in untreated cells were compared. Data showed that the recombinant cells expressed higher levels of *HFE* and *HAMP* mRNA than Wt cells (2.3-fold; p<0.03 and 3.9-fold; p=0.05, respectively) (Figs. 3a and 3b).

**Discussion**

The genes *HFE* and *HAMP* are extremely important for maintaining body iron homeostasis, where the protein HFE modulates *HAMP* induction [1,2] and the induced peptide hepcidin regulates systemic iron homeostasis upon systemic iron elevation [17,18]. However, the intracellular and extracellular iron-sensing mechanisms remain unclear and the upstream responses of the *HFE* mRNA to a range of increasing extracellular iron concentrations and elevated intracellular iron levels, and its co-relation with *HAMP* mRNA have been rarely studied. Therefore, in this short study we aimed to discriminate between the effects of intracellular and extracellular iron-loading. Hence, we examined *HFE* and *HAMP* expressions under high extracellular iron levels, high intracellular iron levels, and simultaneously increased intracellular and extracellular iron levels by treating Wt and recombinant HepG2 cells with/without a range of increasing holo-Tf concentrations. Such studies will not only help in elucidating the iron-sensing mechanisms but also in understanding iron-acquisition in different iron-excess pathologies.

Unlike previous studies where the responses of *HFE* mRNA expression were studied either under a single concentration of holo-Tf [11,19] or in macrophages [19], here, we used a range of holo-Tf concentrations (from 1 g/L to 8 g/L) on HepG2 cells to mimic the gradually elevating extracellular iron loading in different iron-excess pathologies, accounting for the probable different stages of iron loading. Since HFE is the inducer of *HAMP* expression [1,20–22] and hepcidin is a hormone [23], an early response to treatment was expected. Therefore, unlike most previous studies [11–13], here, the duration of treatments was only 6 h. As expected, alterations were observed in both, Wt and recombinant cells at this time-point (Figs. 1b, 1c, 2b and 2c).

***HFE* mRNA expression is responsive to excess extracellular and intracellular iron**

To our knowledge, no study has yet reported the effect of a range of holo-Tf concentrations or saturation on *HFE* mRNA levels. We report for the first time, that elevation in extracellular holo-Tf concentration for 6 h causes elevation in *HFE* mRNA expression in the Wt HepG2 cells (Fig.1b). As this increase occurred in the absence of intracellular iron elevation (Fig.1a), it could be attributed exclusively to the elevated extracellular holo-Tf concentrations, thereby demonstrating the responsiveness of *HFE* mRNA towards excess extracellular iron. Furthermore, high *HFE* mRNA expression in the absence of extracellular iron, but presence of high intracellular iron (as observed in untreated recombinant cells) can be attributed exclusively to the high intracellular iron content (Figs. 3a, 1a and 2a). This indicates the responsiveness of *HFE* mRNA exclusively to high intracellular content. Collectively, *HFE* mRNA expression showed independent sensitivity to extracellular and intracellular iron loading.

In the Wt cells, lack of significant increase in intracellular iron following holo-Tf treatment (Fig. 1a) was anticipated; partly due to the short 6 h duration of treatment, and partly due to the source of iron (holo-Tf) used in this study. Unlike non-transferrin bound iron uptake, in which the pathways of iron acquisition and the corresponding regulatory mechanisms are unclear, transferrin bound iron uptake, as mediated in this study, is a well-understood and well-regulated mechanism. This concept is supported by previous observations in HepG2 cells where Fe-NTA treatment caused a 4-fold increase in cellular iron content compared to an insignificant 1.2-fold increase by 4.5 g/L holo-Tf [11].

Regulated cellular iron uptake follows the principles of iron-response element (IRE)-iron response element binding protein (IRP) system [24,25] that aims at maintaining cellular iron homeostasis. Accordingly, subtle increments in intracellular iron would be sensed by the IRE-replete *TFRC* transcripts and lead to decreased transcript levels and eventually, decreased TfR1 protein expression on cell-surface to prevent further iron-uptake [25]. Such increments would be additionally sensed by the IRE-replete *SLC40A1* (ferroportin) transcripts, lead to increased expression and mediate iron efflux via this iron exporter to maintain intracellular iron homeostasis [24,25]. This is supported by the data (Supplementary Fig.1), which shows that in the Wt HepG2 cells, both, *TFRC* and *SLC40A1* transcripts were downregulated upon holo-Tf treatment, thereby preventing both, iron uptake and iron efflux. Thus, the Wt cells showed no major increase in intracellular iron content upon holo-Tf treatment, demonstrating cellular iron-homeostatic mechanisms in action (Fig. 1a), resembling data from another study [11] and reflecting physiological conditions where excess cellular iron uptake would be prevented under excess extracellular iron to maintain cellular iron homeostasis.

***HAMP* mRNA expression and iron**

In the Wt cells, elevation of *HAMP* expression following holo-Tf supplementation (Fig.1c) is an expected response following an iron stimulus [17,26,27].These elevations occurred in the absence of increased intracellular iron, indicating that an increase in extracellular iron was sufficient for the induction and a major increase in intracellular iron content was not necessary. Interestingly, its wavy pattern of expression over the increasing holo-Tf concentrations displayed a typical hormonal characteristic where increased levels of a stimulant (here, holo-Tf) may not lead to a directly proportional mRNA response. This is because, unlike cytokines, hormone-peptides are ‘premade’ and released from vesicles following a stimulus, like incase of insulin [28]. In the absence of extracellular iron (untreated cells), the high *HAMP* expression in recombinant cells (Fig. 3b), indicated that *HAMP* could be induced exclusively due to high intracellular iron content (Figs. 2a and 1a).

**Interrelationship between *HFE* and *HAMP* expression patterns**

Since HFE is an inducer of *HAMP* expression [1,22], a correlation between the mRNA responses of *HFE* and *HAMP* over the increasing holo-Tf concentrations was envisaged. The Wt cells showed no co-relation between the patterns of their responses (Figs. 1b and 1c), probably reflecting the hormonal characteristic of hepcidin. Conversely, the recombinant cells showed similarities between the patterns of *HFE* and *HAMP* expressions (Figs. 2b and 2c). Data in the recombinant cells showed that under intracellular iron excess, only subtle extracellular iron elevation could increase *HFE* and *HAMP* expressions, while further increase in extracellular iron led to either repression or an unaltered effect (Figs. 2b and 2c). This implies that both these genes can be induced by an external iron stimulus to regulate iron homeostasis, but preferably in the absence of intracellular iron loading, as supported by *HFE* and *HAMP* elevations observed in Wt cells that did not show intracellular iron loading (Fig.1). Accordingly, it could be extrapolated that once intracellular iron loading is attained in iron-excess pathologies such as alcoholic liver disease and hepatitis C infections, the iron-regulatory functionality of *HFE* and *HAMP* is dampened. This could be one of the reasons for deregulated iron metabolism and insufficient hepcidin production in such pathologies that show both, systemic and cellular iron loading along with diminished hepcidin production despite the presence of functional alleles of iron-related genes [5,7,29,30]. Additional experiments and corresponding clinical data will be necessary to provide more evidence to support the resemblance of our findings with such clinical conditions.

Further studies are required to elucidate these mechanisms to better understand the iron-sensing and iron-loading mechanisms; aiming to design therapeutic interventions for iron-excess pathologies other than hemochromatosis.

**Conclusion**

In this short study, the independent effects of extracellular and intracellular iron on *HFE* and *HAMP* expressions were examined. *HFE* mRNA demonstrated independent responsiveness to elevated extracellular and intracellular iron content, suggesting its involvement in sensing both, extracellular and intracellular iron. Under combined intracellular and extracellular iron loading, *HFE* and *HAMP* expressions showed similar patterns and *HAMP* was induced only by low holo-Tf concentration, a scenario resembling certain iron excess pathologies.

**Compliance with Ethical Requirements:** This article does not contain any studies conducted on human or animal subjects.

**Author contributions**

Dr Kosha Mehta: Key researcher; carried out experimental work, performed statistical analyses and wrote the article.

Dr Sebastien Farnaud: Concept of the research.

Dr Vinood B. Patel: Director of studies and final approver for the version of the article to be published.

**Abbreviations**: EMEM: Eagle’s Minimal essential medium; Holo-Tf: holotransferrin; H: hours

**Gene annotations:** *HAMP*, gene encoding hepcidin; *HFE*, gene encoding hemochromatosis protein HFE.

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**Figure Captions and Legends**

**Fig. 1 Effects of holo-Tf supplementation in Wt HepG2 cells**

WtHepG2 cells were treated with holo-Tf for 6 h. Following the treatment, intracellular iron levels were measured and expressed per mg protein (a). *HFE* (b) and *HAMP* (c) mRNA expressions was assessed and expressed relative to untreated (0 g/L) cells. Data is presented as mean ± SEM (n=3). \*p<=0.05 compared to untreated (0 g/L) controls.

**Fig. 2Effects of holo-Tf supplementation in rec-TfR1 HepG2 cells**

Rec-TfR1 HepG2 cells were treated with holo-Tf for 6 h. Following the treatment, intracellular iron levels were measured and expressed per mg protein (a). *HFE* (b) and *HAMP* (c) mRNA expressions was assessed and expressed relative to untreated (0 g/L) cells. Data is presented as mean ± SEM (n=3). \*p<0.03, \*\*p<0.01 and #p=0.07 compared to untreated (0 g/L) controls. ^p<0.03 compared to 2 g/L treatment.

**Fig. 3*HFE and HAMP* mRNA expressions in rec-TfR1 HepG2 cells relative to WtHepG2 cells**

The mRNA expressions of *HFE* (a) and *HAMP* (b) in the recombinant cells were expressed relative to WtHepG2 cells under untreated conditions at the 6 h time point.Data is presented as mean ± SEM (n=3). \*p<=0.05 compared toWtHepG2 cells.

**Table 1 Summary of *HFE* and *HAMP* expression patterns**

|  |  |  |
| --- | --- | --- |
|  | ***HFE* mRNA expression**  | ***HAMP* mRNA expression**  |
|  |  **Wt cells**  | **Recombinant cells** | **Wt cells** | **Recombinant cells** |
| **1g/L** | **-** |  |  |  |
| **2 g/L** | **-** |  | **-** |  |
| **5 g/L** |  | **-** |  | **-** |
| **8 g/L**  |  | **N/A** |  | **N/A** |

Key to Table 1:

**-** : unaltered expression

 : Increased expression

 : decreased expression

N/A: not applicable as the expression was not studied.

**Fig. 1a**

**Fig. 1b**

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**Fig. 1c**

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**Fig. 2a**

**Fig. 2b**

**Fig. 2c**

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**Fig. 3a**

**Fig. 3b**

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Supplementary Fig.1.

a) b)

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**Supplementary Fig.1 mRNA expression of iron-uptake and iron-efflux genes in Wt HepG2 cells**

Holo-Tf-induced mRNA expression of (a) *TFRC* (encoding transferrin receptor 1) and (b) *SLC40A1* (encoding ferroportin) in the Wt HepG2 cells have been shown. \*p<=0.05 compared to untreated (0 g/L) controls. Data is presented as mean ± SEM (n=3).